



## Popular Article

Domain: Agriculture Science

Vol 4 Issue 12, December 2025, 6086-6089

# Evaluating The Efficacy of Bavistin Against *Rhizoctonia* sp., *Helminthosporium* sp., & *Alternaria* sp.

Sagnik Ghosh<sup>1</sup>, Ishanu Mandal\*<sup>1</sup>

<sup>1</sup>Faculty of Agriculture, JIS University, Agarpara, Kolkata: 700109, W.B, India.

\*Corresponding Email: [ishanumandal80@gmail.com](mailto:ishanumandal80@gmail.com)

DOI:10.5281/TrendsInAgri.18155866

### Abstract

This study assesses the fungicidal efficacy of Bavistin (Carbendazim 50% WP) against three significant pathogens—*Rhizoctonia* sp., *Helminthosporium* sp., and *Alternaria* sp.—via the poison food method. Depending on the concentration, Bavistin inhibited fungal growth in varying degrees, with LD<sub>50</sub> values determined as 68 ppm, 648 ppm, and 71 ppm respectively. These findings underscore the potential of Bavistin as a cost-effective, targeted solution to fungal infestations, offering economic and ecological benefits for Indian farmers, particularly in West Bengal. Optimizing dosage can address resistance and environmental impact concerns while enhancing crop yield.

**Keywords:** Bavistin, *Rhizoctonia*, *Helminthosporium*, *Alternaria*, Poison Food Method

### INTRODUCTION

Fungal diseases are a major challenge in agriculture, causing significant crop losses worldwide. In India, pathogens such as *Rhizoctonia* sp. (Boerema & Hamers, 1988), *Helminthosporium* sp. (Thuy *et al.*, 2023), and *Alternaria* sp. (Simmons, 2000) affect essential crops like rice, wheat, and vegetables, critical for food security and the economy, especially in regions like West Bengal. The chemical fungicide Bavistin, a carbendazim-based formulation, has been widely used to mitigate fungal infections (Kumar & Reddy, 2021). However, unregulated usage often leads to resistance, environmental concerns, and financial strain on farmers. This study employs the poison food technique (Nene & Thapliyal, 1979) to quantify Bavistin's efficacy and establish optimal dosages for effective pathogen control.

### MATERIALS AND METHODS

#### Experimental Setup

- **Pathogens Tested:** Cultures of *Rhizoctonia* sp., *Helminthosporium* sp., and *Alternaria* sp. (7 days old).
- **Fungicide:** Bavistin (Carbendazim 50% WP) at varying concentrations.

- Media Preparation:** The Potato Dextrose Agar (PDA) medium was prepared using boiled potato extract, dextrose, and agar-agar powder.
- Concentrations:** Working solutions of Bavistin were prepared at 50, 100, 250, 500, and 1000 ppm, alongside a control group with no fungicide.

### Poison Food Method

- Sterile PDA media were infused with respective fungicide concentrations.
- Mycelial discs (6 mm diameter) were placed at the centre of PDA plates.
- Plates were incubated at  $25 \pm 1^\circ\text{C}$  for three days in a controlled environment.
- Radial mycelial growth was measured, and percentage inhibition was calculated using the formula:

$$\text{Percent Growth Inhibition}(\%) = \frac{\text{Diameter in Control} - \text{Diameter in Treatment}}{\text{Diameter in Control}} \times 100$$

### LD<sub>50</sub> Determination

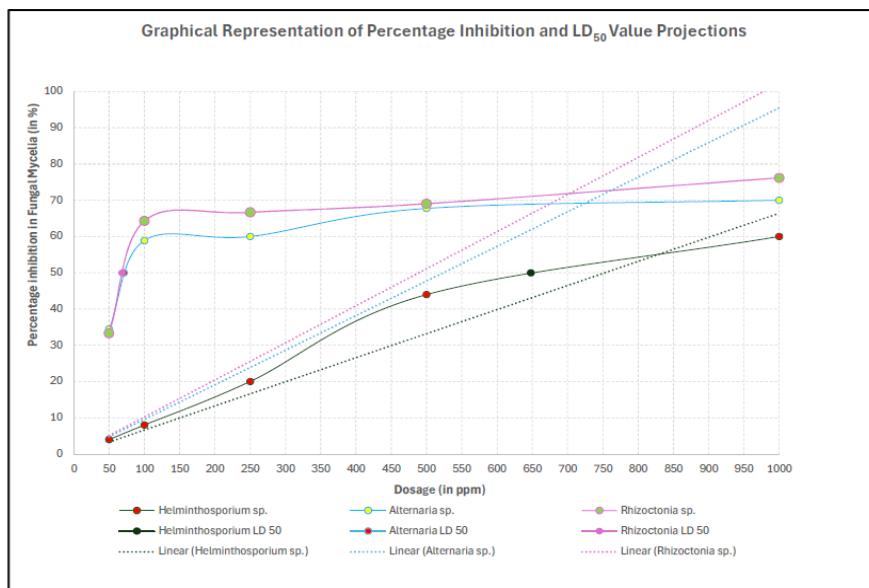
The LD<sub>50</sub> value for each pathogen was graphically estimated by plotting inhibition percentages against fungicide concentrations. The value corresponds to the concentration inhibiting 50% growth (Nene & Thapliyal, 1979).

### CALCULATION

Dose (in ppm)	Diameter of Different Mycelial Growth (in cm)			Percentage Inhibition in Fungal Mycelia (%)		
	<i>Rhizoctonia</i> sp.	<i>Helminthosporium</i> sp.	<i>Alternaria</i> sp.	<i>Rhizoctonia</i> sp.	<i>Helminthosporium</i> sp.	<i>Alternaria</i> sp.
Control	4.2	2.5	9	0	0	0
50	2.8	2.4	5.9	$\approx 33.33$	$\approx 4$	$\approx 34.44$
100	1.5	2.3	3.7	$\approx 64.28$	$\approx 8$	$\approx 58.89$
250	1.4	2	3.6	$\approx 66.67$	$\approx 20$	$\approx 60$
500	1.3	1.4	2.9	$\approx 69.05$	$\approx 44$	$\approx 67.78$
1000	1	1	2.7	$\approx 76.19$	$\approx 60$	$\approx 70$

**Table 1:** Tabulation of the Observed Mycelial Growth Rate of three pathogens and per cent inhibition calculations (As per the aforesaid formula) at different doses of Bavistin application.

## COMPUTATION



**Figure 1:** Graphical computation of the data calculated in Table 1 via MS Excel 365/Microsoft 365 and observation of the LD<sub>50</sub> Values for each pathogen.

Name of the pathogen	Ordinate (y-coordinate)	Axis (x-coordinate/LD <sub>50</sub> Values)
<i>Rhizoctonia</i> sp.	50	≈ 68
<i>Helminthosporium</i> sp.	50	≈ 648
<i>Alternaria</i> sp.	50	≈ 71

**Table 2:** Computation of the LD<sub>50</sub> values for all the pathogens observed from **Figure 1**.

## RESULTS AND DISCUSSION

**Growth Inhibition Trends:** *Rhizoctonia* sp. showed increasing inhibition with concentration, reaching 76.19% at 1000 ppm, while *Helminthosporium* sp. was less sensitive, achieving a maximum inhibition of 60% at 1000 ppm and *Alternaria* sp. exhibited 70% inhibition at the highest concentration.

LD<sub>50</sub> Values:

- *Rhizoctonia* sp.: 68 ppm
- *Helminthosporium* sp.: 648 ppm
- *Alternaria* sp.: 71 ppm

## Discussion

Bavistin demonstrated high efficacy against *Rhizoctonia* sp. and *Alternaria* sp., with relatively low LD<sub>50</sub> values indicating effectiveness at lower concentrations. However, *Helminthosporium* sp. required significantly higher doses for comparable inhibition, suggesting intrinsic resistance mechanisms. This highlights the need for tailored fungicide applications based on pathogen-specific sensitivity to prevent overuse and resistance.

## Relevance to West Bengal Agriculture

West Bengal, a region heavily reliant on crops like rice, jute, and vegetables, suffers substantial economic losses due to fungal infections. Using Bavistin at optimal concentrations can reduce crop losses by targeting specific pathogens and enhance productivity, directly benefiting small-scale farmers (Kumar & Reddy, 2021).

## CONCLUSION

This study confirms Bavistin's utility as an effective fungicide for managing *Rhizoctonia sp.*, *Helminthosporium sp.*, and *Alternaria sp.* in controlled settings. Adopting precise application strategies based on LD<sub>50</sub> values can significantly improve its field efficacy, offering a sustainable solution for combating fungal diseases in Indian agriculture. Further field trials are recommended to validate laboratory findings under real-world conditions.

## REFERENCES

Boerema, G. H., & Hamers, M. E. (1988). Check-list for scientific names of common parasitic fungi. Series 3a: Fungi on bulbs: Liliaceae. *Netherlands Journal of Plant Pathology*, 94, 1-29. <https://doi.org/10.1007/BF02001882>.

Kumar, P. & Reddy C. N. (2021). In Vitro Toxicity of Bavistin (Carbendazim 50% WP) On *Sclerotium Rolfsii* Sacc. *International Journal of Advance Research, Ideas and Innovations in Technology*. 7(1), 135-137. ISSN: 2454-132X.

Nene, Y.L. & Thapliyal, P.N. (1979) *Fungicides in Plant Disease Control*. 2nd Edition, Oxford and IBH Pub. Co., New Delhi. ISBN 10: 9386479850.

Simmons, E. G. (2000). *Alternaria* themes and variations (244-286) species on Solanaceae. *Mycotaxon*, 75, 1-115. ISSN (Print): 0093-4666.

Thuy, T. T. T., Lübeck, M., Smedegaard-Petersen, V., de Neergaard, E., & Jørgensen, H. J. L. (2023). Infection Biology of *Bipolaris oryzae* in Rice and Defence Responses in Compatible and Less Compatible Interactions. *Agronomy*, 13(1), 231. <https://doi.org/10.3390/agronomy13010231>.